

Circular RNAs in the pathogenesis of sepsis and their clinical implications: A narrative review

Lin Wei ^{*1}*MMed*, Yongpeng Yang ^{*1}*MBBS*, Weikai Wang ¹*MMed*, Ruifeng Xu ¹*MMed*

ABSTRACT

Introduction: Sepsis is defined as a life-threatening complication that occurs when the body responds to an infection attacking the host. Sepsis rapidly progresses and patients deteriorate and develop septic shock, with multiple organ failure, if not promptly treated. Currently no effective therapy is available for sepsis; therefore, early diagnosis is crucial to decrease the high mortality rate. Genome-wide expression analyses of patients in critical conditions have confirmed that the expression levels of the majority of genes are changed, suggesting that the molecular basis of sepsis is at the gene level. This review aims to elucidate the role of circular (circ) RNAs in the pathogenesis of sepsis and sepsis-induced organ damage. In addition, the feasibility of using circRNAs as novel diagnostic biomarkers for sepsis is also discussed, as well as circRNA-based therapy.

Method: This narrative review is based on a literature search using Medline database. Search terms used were “circular RNAs and sepsis”, “circRNAs and sepsis”, “non-coding RNAs and sepsis”, “ncRNAs and sepsis”, “circRNAs and septic pathogenesis”, “circRNAs and septic model”, “circRNAs and septic shock” and “circRNAs, biomarker, and sepsis”.

Results: Numerous studies indicate that circRNAs might exert pivotal roles in regulating the immune system of the host against various pathogens, such as bacteria and viruses. Dysregulation of circRNA expression levels has been confirmed as an early event in sepsis and associated with the inflammatory response, immunosuppression and coagulation dysfunction. This impairment in regulation eventually leads to multiple organ dysfunctions, including of the kidneys, lungs and heart.

Conclusion: By investigating the regulation of circRNAs in sepsis, new molecular targets for the diagnosis and intervention of sepsis can be identified. Such an understanding will be important for the development of therapeutic drugs.

Ann Acad Med Singap 2022;51:221-7

Keywords: Acute kidney injury, biomarker, circRNAs, inflammation, sepsis

INTRODUCTION

Sepsis is a condition with life-threatening organ dysfunction, resulting from abnormal responses of the host to various infections.¹ The underlying pathogenic mechanisms include an imbalanced inflammatory response, immune disorder, neuroendocrine abnormality, coagulopathy, mitochondrial damage and endoplasmic reticulum stress.² A recent study reported that the age-standardised sepsis incidence rate fell by 37.0% and the mortality rate decreased by 52.8% from 1990 to 2017.³ Despite the declining incidence and mortality

rates, sepsis is still a global problem, with the highest health-related burden in sub-Saharan Africa.³

Biomarkers can be used in clinics to evaluate the pathophysiological process of various diseases, and play important roles in assisting the diagnosis, monitoring the efficacy of treatment and influencing the prognosis. There have been >170 biomarkers identified that are associated with sepsis; however, the majority of them lack sensitivity or specificity, and only a few have been used in the clinical diagnosis of infections, including C-reactive protein, procalcitonin, high mobility group protein B-1,

¹ Center for Children's Intensive Care, Gansu Provincial Maternity and Child-care Hospital, Lanzhou, China

Correspondence: Dr Ruifeng Xu, Center for Children's Intensive Care, Gansu Provincial Maternity and Child-care Hospital, No.143 Qilihe North Street, Qilihe District, Lanzhou, Gansu 730050, China.

Email: xurf110gsfy@sina.com

* Contributed equally to this work

CLINICAL IMPACT

What is New

- This review provides an update on the understanding of circular RNAs (circRNAs) in the pathogenesis of sepsis and the associated organ dysfunction. It investigates their feasibility as effective and efficient diagnostic and therapeutic biomarkers for sepsis.

Clinical Implications

- The understanding of circRNA-modulated sepsis-induced organ failure is still at a very early stage. Thus, further research is urgently required to investigate the regulation of circRNAs in the progression of different diseases, and to identify specific circRNAs in certain diseases.

interleukins and soluble triggering receptor expressed on myeloid cells-1.⁴ Therefore, the severity and recovery of sepsis cannot be objectively evaluated, which limits the wide application of these biomarkers in a clinical setting. Thus, identification of more effective biomarkers is an urgent need to improve diagnosis/prognosis.

Circular (circ) RNAs are newly identified endogenous non-coding RNAs formed by exon scrambling during the splicing process. They are typically covalently closed-loop molecules, which distinguish them from the other 2 linear non-coding RNAs—microRNAs (miRNA/miR) and long non-coding RNAs—which possess caps at the 5' terminal and tails at 3' terminal.⁵ With rapid evolution of high-throughput sequencing techniques, circRNAs have been found to be widely distributed in various eukaryotes. They exert significant roles in gene transcription and participate in a range of cellular events such as cell differentiation, apoptosis, autophagy and proliferation, which are all associated with septic pathogenesis.⁶ Here, we provide an update on the progress of our understanding of circRNAs in the pathogenesis of sepsis and the associated organ dysfunction. We also investigate their feasibility as effective and efficient diagnostic and therapeutic biomarkers for sepsis.

METHOD

This narrative review is based on a literature search of the Medline database. Search terms used were “circular RNAs and sepsis”, “circRNAs and sepsis”, “non-coding RNAs and sepsis”, “ncRNAs and sepsis”, “circRNAs and septic pathogenesis”, “circRNAs and septic model”, “circRNAs and septic shock” and “circRNAs, biomarker, and sepsis”.

Overview of circRNAs

circRNAs have a stable structure, are highly expressed, have specific tissue distribution patterns and are highly conserved among different species.⁷ They are widespread and substantial within transcriptomes, an observation confirmed using high-throughput sequencing and analysis platforms.⁸ circRNAs are principally derived from exons of protein-coding genes and generally formed via the cyclisation of special mRNAs by reverse splicing, which produces covalently closed-loop structures with a length of about 100 nucleotides.⁹ circRNAs are generated by multiple mechanisms, including cyclisation via lariat, intron pairing or RNA binding protein pairing. circRNAs cannot be cut by RNA exonuclease.¹⁰ In addition, based on their origin of biogenesis, circRNAs are categorised into exon-, intron-, exon-intron-, and intergenic circRNAs.¹¹

Many circRNAs have been indicated to function as “miRNA sponges” to regulate miRNAs in various body fluids and to also function as protein sponges to determine the concentration of proteins in cells.⁵ circRNAs have a longer half-life than their homologues and show good stability.¹² It is well-known that circRNAs are not simply the by-products from mis-splicing; instead they are actively involved in pathological processes of diverse diseases.¹³ miRNAs have been found to be differentially expressed during the development of sepsis, suggesting certain circRNAs might also be involved. For example, circ_0091702 was found to serve as a sponge for miR-545-3p to alleviate sepsis-induced acute kidney injury (AKI) by increasing the expression level of thrombospondin 2.¹⁴ These biological characteristics and physiological functions suggest that circRNAs have the potential to be septic biomarkers.

In addition, as miRNA sponges, circRNAs also function as a “miR reservoir”. For example, circ-HIAT1 was found to target miR-29a-3p and miR-29c-3p, and increase the stability of miRNAs in human atherosclerosis and cancer.¹⁵ In solid tumours, circ-HIAT1 was found to target matrix metalloproteinases that were increased in both the serum and lung tissues in patients with severe sepsis.¹⁶ The high serum level of miR-29a-3p secreted by immune cells has shown a prognostic value in evaluating the 28-day mortality rate in patients with sepsis, while circ-MYLK and circ-CTDP1 have shown a modulatory role in the expression levels of miR-29a-3p.¹⁷

circRNAs in sepsis-related inflammation

miRNAs have been confirmed to regulate the cytokine storm of sepsis. For example, miRNAs regulated the differential expression of many key cytokines that were involved in sepsis, including TNF- α , IL-6, IL-10 and IL-18.¹⁸ circRNAs have been proposed to exert essential

roles during different stages of sepsis by regulating lipopolysaccharide (LPS)-induced inflammation and the activation of NF- κ B signalling via sponging miRNAs. A recent study found that knockdown of circ_0114428 expression inhibited cereblon expression, and attenuated LPS-induced inflammation and oxidative stress in human kidney 2 (HK2) cells by sponging miR-495-3p.¹⁹ Xiong et al. found that the knockdown of circ_0003420 expression could attenuate the effect of LPS on cell apoptosis, proliferation and inflammation by targeting NPAS4 mRNA.²⁰ Wei et al. showed that overexpression of hsa_circ_0068,888 could suppress the LPS-induced inflammatory response and oxidative stress, while knockdown of expression could increase these processes.²¹ A further study found that hsa_circ_0068,888 inhibited the activation of NF- κ B signalling by sponging miR-21-5p.²¹ Furthermore, Liu et al. found that circ_0001105 protected the integrity of the intestinal barrier from intestinal inflammation and oxidative stress in septic rats, providing a new perspective to treat sepsis.²²

circRNAs in sepsis-related immunosuppression

The majority of patients with sepsis may die during the early stage of the cytokine storm. Patients who survive this stage can exhibit immunosuppression—they fail to eliminate primary infections, develop secondary opportunistic infections, and viruses can potentially reactivate—which seriously affects their survival.² circMAN2B2, a circRNA abundant in glioma tissues, was found to regulate S100A8 expression levels by inhibiting miR-1205.²³ S100A8 is known to be an important modulator involved in the immunosuppression of sepsis,²⁴ suggesting a potential role of circMAN2B2 in sepsis-related immunosuppression. miRNAs are able to suppress ZEB1/2-mediated drug resistance and immunosuppression; therefore, circRNAs as upstream modulators of the miR/ZEB1 axis could have a possible role in sepsis-induced immunosuppression.²⁵ circMET was found to drive immunosuppression in hepatic carcinoma via the miR-30-5p/zinc finger protein SNAI1 (Snail) axis,²⁶ while Snail was markedly elevated in glomerular tissue in septic patients,²⁷ suggesting potential roles of circMET's in sepsis-related immunosuppression.

circRNAs in sepsis-related coagulation dysfunction

Activation of the coagulation system is affected during sepsis, which is a critical indicator during the development of sepsis. Endothelial cells (ECs) exert an essential effect on maintaining vascular homeostasis and are the primary targets of inflammatory mediators in sepsis. The persistent damage to ECs could cause organ failure.²⁸ A recent study showed that the overexpression of circ-C3P1 suppressed cell apoptosis and pro-inflammatory cytokines

in pulmonary microvascular ECs in an LPS-induced sepsis mouse model by negatively modulating miR-21.²⁹ These findings indicate that the endothelial function could be damaged by the abnormal expression level of circRNAs, which finally leads to coagulation disorder and expedited septic progression.

circRNAs and sepsis-related organ dysfunction

circRNAs in sepsis-induced AKI

AKI is a frequently observed condition in the clinic with a high incidence rate, and acute inflammation and tissue injury are common indications. Sepsis is one of the most common causes of AKI, accounting for more than half of AKI cases. Among them, sepsis, triggered by LPS, is the dominant factor of AKI in patients in a critical condition, and is often used to establish in vitro sepsis-induced AKI models. Recently, a range of circRNAs have been shown to be involved in the development of AKI. For example, knockdown of circ-FANCA was found to alleviate LPS-induced HK2 cell injury by modulating the miR-93-5p/OXSR1 axis in sepsis-induced AKI.³⁰ circ-Ttc3 was found to reduce the inflammatory response and oxidation by targeting the miR-148a/Rcan2 axis in rats with sepsis-induced AKI, indicating that circ-Ttc3 could be a potential therapeutic target.³¹ In addition, the involvement of circRNAs in regulating programmed cell death and cell cycle progression indicates that they are novel modulators for sepsis-induced AKI.²⁰ For example, circRar1 was found to induce the transcriptional activity of apoptosis-related factors in lead-induced neurotoxicity by regulating miR-671.³²

circRNAs in sepsis-related acute respiratory distress syndrome

Acute respiratory distress syndrome (ARDS) is known as an independent risk factor for mortality in patients with sepsis. Accumulating evidence has shown that LPS can induce the activation and migration of the monocyte-macrophage system. This system further engulfs inflammatory particles invading the lungs, and secretes a mass of bioactive substances that facilitate neutrophil migration during ARDS development.³³ A recent study investigated the circ-ANKRD36-associated molecular mechanism underlying sepsis-related ARDS and confirmed that circ-ANKRD36 expression levels were markedly increased in the serum of patients with sepsis-related ARDS. In addition, knockdown of circ-ANKRD36 expression reduced cell viability and migration in LPS-treated RAW264.7 cells by sponging miR-330, suggesting a novel strategy in treating sepsis-related ARDS.³⁴ Another study confirmed the protective roles of the P2X7R antagonist in mice with sepsis-related

ARDS by regulating the expression levels of circ_0001679 and circ_0001212, suggesting these 2 circRNAs could be potential targets for sepsis-related ARDS treatment.³⁵

circRNAs in sepsis-related myocardial dysfunction

Sepsis-related myocardial dysfunction (SIMD) is another significant complication following sepsis. Myocardial depressant factors, apoptosis, inflammatory cytokines, complement activation as well as nitric oxide have been found to contribute to the pathological course of SIMD.³⁶ The current strategies to treat SIMD include maintaining the stability of haemodynamics and supporting the normal cardiac function. Nevertheless, specific medications were limited, due to the undefined regulatory mechanism underlying the pathogenesis of SIMD. circRNAs are emerging as important modulators in a range of biophysiological processes, including myocardial dysfunction. For example, circ-HIPK2 was found to regulate proliferation and differentiation during myogenesis by targeting ribosomal protein Rpl7.³⁷ circACSL1 was found to modulate MAPK14 expression by sponging miR-8055 and aggravating myocardial inflammation and damage. This effect suggests that circACSL1 could be an effective biomarker used in the diagnosis and treatment of myocardial dysfunction.³⁸ In addition, miR-23b-induced activation of myocardial fibrosis has been confirmed as a key factor of myocardial dysfunction in advanced sepsis, while a miR-23b inhibitor was found to reduce cardiac fibrosis during the late stage of sepsis. A recent study found that the inhibition of miR-23b by circ_0005075 prevented polymicrobial sepsis-induced cardiac disorder by modulating TG-interacting factor 1 (TGIF1), phosphatase and tensin homologue deleted on chromosome 10 (PTEN), suggesting that circ_0005075 might be an effective modulator in the treatment of SIMD.³⁹ Recent studies on the roles of circRNAs in sepsis-induced organ failure are shown in Table 1.

circRNAs in virus-induced sepsis

Sepsis is commonly induced by bacterial infection; however, it can also be triggered by viral or fungal infection, causing weaker inflammatory responses. Until now, there is no definite diagnostic criterion for virus-induced sepsis. The discovery of circRNAs shows their advantages over other biomarkers, such as a high stability in the presence of viral infection, and may thus be used as potential diagnostic tools. A recent study found that NF90/NF110 released from host circRNP complexes could couple with viral mRNAs as part of the anti-viral immune response and against viral infection.⁵⁵ As circRNA expression is generally low, a certain group of

circRNAs, not a single specific circRNA, may function together as molecular reservoirs of NF90/NF110 for rapid immune response upon viral infection.⁵⁵

Technologies used to identify and evaluate circRNAs as biomarkers

The difficulty in distinguishing circRNAs from other non-coding RNAs has led to the late discovery of circRNAs. Currently, subject to the limited detection methods, circRNAs have to lose its circularity in order to be successfully detected. A protocol called Circle-Sequencing (Circle-Seq) has been recently proposed and used to process linear RNAs using RNase R enzyme, while keeping circRNAs intact.⁵⁶ However, this protocol shows certain limitation in determining the circularity of a RNA transcript as certain circRNAs are sensitive to RNase R, and might lead to false negative results. Other strategies, including 2-dimensional denaturing polyacrylamide gel electrophoresis, ribosomal RNA or poly(A) depletion, are also used to isolate and concentrate circRNAs in samples with unknown efficacy in clinical practice.⁵⁷ Microarray is another promising diagnostic tool, which can be used to determine the relative levels of different circRNAs due to its sensitivity and specificity. However, it is only able to evaluate those circRNAs covered in the array, and cannot be used to measure the total amount of circRNAs.⁵⁸

Currently, reverse transcription-quantitative polymerase chain reaction is broadly utilised to recognise and quantify circRNAs. It shows great advantages over other methods as it is simple and inexpensive as a detection tool for diagnostic biomarkers.⁵⁹ In addition, RNA-sequencing (RNA-Seq) technology, combined with bioinformatics analysis, enables the comprehensive study on circRNAs, and significantly contributes to the discovery and characterisation of circRNAs.⁶⁰ Disease-related circRNAs have been identified in human peripheral blood using RNA-Seq.⁴⁵

circRNAs have a crucial role in sepsis, thus it is essential to clarify the underlying mechanisms of different circRNAs involved in the pathogenesis of sepsis, in addition to identifying novel circRNAs that regulate the heterogeneity of sepsis.

CONCLUSION

Diagnosis of early sepsis is extremely important to maximise the survival rate in patients with sepsis. The availability of accurate biomarkers will be particularly beneficial to enable the delivery of prompt and appropriate treatment. However, none of the current biomarkers, that are clinically evaluated could offer 100% specificity for the diagnosis of sepsis.⁴ circRNAs, as newly identified

Table 1. Recent studies on the roles of circRNAs in sepsis-induced organ failure

circRNAs	Downstream targets	Functions
circ-ANKRD36	miR-330	Its knockdown suppresses cell viability and migration in LPS-stimulated cells ³⁴
circ-BNIP3L	miR-370-3p/MYD88	Its knockdown alleviates LPS-induced renal tubular epithelial cell injury ⁴⁰
circ-C3P1	miR-21	Attenuates pro-inflammatory cytokine production and cell apoptosis in septic ALI ²⁹
circ-DMNT3B	miR-20b-5p	Its downregulation is conducive to intestinal mucosal permeability dysfunction ⁴¹
circ-FADS2	miR-15a-5p	Suppresses LPS-induced lung cell apoptosis ⁴²
circ-FANCA	miR-93-5p/OXSR1	Its knockdown alleviates LPS-induced HK2 cell injury ³⁰
circ-Fryl	miR-490-3p/SIRT3	Attenuates sepsis-induced lung injury ⁴³
circ-HIPK3	miR-29b	Promotes homeostasis of the intestinal epithelium ⁴⁴
circ-PRKCI	miR-545	Associated with sepsis risk, disease severity and 28-day mortality ⁴⁵
	miR-545/ZEB2	Relieves LPS-induced HK2 cell injury ⁴⁶
	miR-106b-5p/GAB1	Alleviates LPS-induced HK2 cell injury ⁴⁷
circ-PTK2	miR-181c-5p/HMGB1	Regulates microglia activation and hippocampal neuronal apoptosis induced by sepsis ⁴⁸
circ-TLK1	miR-106a-5p/HMGB1	Promotes sepsis-associated AKI by regulating inflammation and oxidative stress ⁴⁹
circ-Ttc3	miR-148a/Rcan2	Regulates inflammation and oxidative stress in septic rats with AKI ³¹
circ-VMA21	miR-199a-5p/NRP1	Ameliorates LPS-induced AKI ⁵⁰
	miR-9-3p/SMG1	Ameliorates sepsis-associated AKI inflammation and oxidative stress ⁵¹
circ_0001105	YAP1	Protects intestinal barrier in septic rats by inhibiting inflammation and oxidative damage ²²
circ_0003420	NPAS4	Mediates inflammation in sepsis-induced liver damage ²⁰
circ_0068,888	miR-21-5p	Protects against LPS-induced HK2 cell injury ²¹
circ_0091702	miR-545-3p/THBS2	Attenuates sepsis-related AKI ⁵²
	miR-182/PDE7A	Relieves LPS-induced cell injury ⁵³
circ_0114428	miR-495-3p/CRBN	Regulates sepsis-induced kidney injury ¹⁹
circ_104484/circ_104670	-	Serves as potential novel biomarkers and therapeutic targets for sepsis ⁵⁴

AKI: acute kidney injury; ALI: acute lung injury; circ/circRNA: circular RNA; CRBN: cereblon; GAB1: growth factor receptor binding 2-associated binding protein 1; HK2: human kidney 2; HMGB1: high mobility group box 1; LPS: lipopolysaccharide; miR: microRNA; MYD88: myeloid differentiation primary response protein 88; NPAS4: neuronal PAS domain protein 4; NRP1: neuropilin-1; OXSR1: oxidative stress-responsive 1; PDE7A: phosphodiesterase 7A; Rcan2: regulator of calcineurin 2; SIRT3: sirtuin 3; SMG1: suppressor of morphogenesis in genitalia 1; THBS2: thrombospondin 2; YAP1: Yes-associated protein 1; ZEB2: zinc finger E-box binding homeobox 2

Superscript numbers: Refer to numbers in REFERENCES

non-coding RNAs, have been associated with various molecular responses such as inflammation, immune suppression and endothelial function, and are significantly altered during the progression of sepsis.

circRNAs show promising potential as biomarkers based on their strong resistance to exonucleases and high stability in blood. Their levels are increased and have an average half-life of 48 hours compared to 10 hours for other linear non-coding RNAs in the serum.⁶¹ The putative function of circRNAs as miRNA sponges makes them particularly interesting therapeutic targets for future research. Furthermore, circRNAs have tissue-

specific and developmental stage-specific features. Their expression levels have been shown to be associated with the occurrence and development of various diseases, including sepsis. However, the understanding of circRNA-modulated sepsis-induced organ failure is still at a very early stage. Further research is urgently required to investigate the regulation of circRNAs in the progression of different diseases, and to identify specific circRNAs in certain diseases. Therefore, circRNAs are attracting increasing attention in sepsis research, and may play significant roles in the diagnosis, treatment and prognosis of sepsis.

REFERENCES

1. Singer M, Deutschman CS, Seymour CW, et al. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016;315:801-10.
2. Huang M, Cai S, Su J. The Pathogenesis of Sepsis and Potential Therapeutic Targets. *Int J Mol Sci* 2019;20:5376.
3. Rudd KE, Johnson SC, Agesa KM, et al. Global, regional, and national sepsis incidence and mortality, 1990-2017: analysis for the Global Burden of Disease Study. *Lancet* 2020;395:200-11.
4. Nikolaos T, Anastasia K, Stylianos E, Biomarkers in infection and sepsis: Can they really indicate final outcome? *Int J Antimicrob Agents* 2015;46(Suppl 1):S29-32.
5. Li X, Yang L, Chen LL. The Biogenesis, Functions, and Challenges of Circular RNAs. *Mol Cell* 2018;71:428-42.
6. Zhang TN, Li D, Xia J, et al. Non-coding RNA: a potential biomarker and therapeutic target for sepsis. *Oncotarget* 2017;8:91765-78.
7. Barrett SP, Salzman J. Circular RNAs: analysis, expression and potential functions. *Development* 2016;143:1838-47.
8. Jeck WR, Sorrentino JA, Wang K, et al. Circular RNAs are abundant, conserved, and associated with ALU repeats. *RNA* 2013;19:141-57.
9. Jeck WR, Sharpless NE. Detecting and characterizing circular RNAs. *Nat Biotechnol* 2014;32:453-61.
10. Ashwal-Fluss R, Meyer M, Pamudurti NR, et al. CircRNA biogenesis competes with pre-mRNA splicing. *Mol Cell* 2014;56:55-66.
11. Huang C, Shan G. What happens at or after transcription: Insights into circRNA biogenesis and function. *Transcription* 2015;6:61-4.
12. Euka Y, Lauriola M, Feldman ME, et al. Circular RNAs are long-lived and display only minimal early alterations in response to a growth factor. *Nucleic Acids Res* 2016;44:1370-83.
13. Qi L, Yan Y, Chen B, et al. Research progress of circRNA as a biomarker of sepsis: a narrative review. *Ann Transl Med* 2021;9:720.
14. Tan M, Bei R. Circ_0091702 serves as a sponge of miR-545-3p to attenuate sepsis-related acute kidney injury by upregulating THBS2. *J Mol Histol* 2021;52:717-28.
15. Holdt LM, Stahringer A, Sass K, et al. Circular non-coding RNA ANRIL modulates ribosomal RNA maturation and atherosclerosis in humans. *Nat Commun* 2016;7:12429.
16. Geng Y, Jiang J, Wu C. Function and clinical significance of circRNAs in solid tumors. *J Hematol Oncol* 2018;11:98.
17. Huo R, Dai M, Fan Y, et al. [Predictive value of miRNA-29a and miRNA-10a-5p for 28-day mortality in patients with sepsis-induced acute kidney injury.] (Article in Chinese) *Nan Fang Yi Ke Da Xue Xue Bao* 2017;37:646-51.
18. Beltrán-García J, Osca-Verdegal R, Nacher-Sendra E, et al. Circular RNAs in Sepsis: Biogenesis, Function, and Clinical Significance. *Cells* 2020;9:1544.
19. He Y, Sun Y, Peng J. Circ_0114428 Regulates Sepsis-Induced Kidney Injury by Targeting the miR-495-3p/CRBN Axis. *Inflammation* 2021;44:1464-77.
20. Xiong H, Wang H, Yu Q. Circular RNA circ_0003420 mediates inflammation in sepsis-induced liver damage by downregulating neuronal PAS domain protein 4. *Immunopharmacol Immunotoxicol* 2021;43:271-82.
21. Wei W, Yao Y, Bi H, et al. Circular RNA circ_0068,888 protects against lipopolysaccharide-induced HK-2 cell injury via sponging microRNA-21-5p. *Biochem Biophys Res Commun* 2021;540:1-7.
22. Liu S, Zhang D, Liu Y, et al. Circular RNA circ_0001105 protects the intestinal barrier of septic rats by inhibiting inflammation and oxidative damage and YAP1 expression. *Gene* 2020;755:144897.
23. Xiong J, Wang T, Tang H, et al. Circular RNA circMAN2B2 facilitates glioma progression by regulating the miR-1205/S100A8 axis. *J Cell Physiol* 2019;234:22996-3004.
24. Dubois C, Marcé D, Faivre V, et al. High plasma level of S100A8/S100A9 and S100A12 at admission indicates a higher risk of death in septic shock patients. *Sci Rep* 2019;9:15660.
25. Ashrafzadeh M, Ang HL, Moghadam ER, et al. MicroRNAs and Their Influence on the ZEB Family: Mechanistic Aspects and Therapeutic Applications in Cancer Therapy. *Biomolecules* 2020;10:1040.
26. Huang XY, Zhang PF, Wei CY, et al. Circular RNA circMET drives immunosuppression and anti-PD1 therapy resistance in hepatocellular carcinoma via the miR-30-5p/snail/DPP4 axis. *Mol Cancer* 2020;19:92.
27. Wang S, Wang J, Zhang Z, et al. Decreased miR-128 and increased miR-21 synergistically cause podocyte injury in sepsis. *J Nephrol* 2017;30:543-50.
28. Ito T, Kakuuchi M, Maruyama I. Endotheliopathy in septic conditions: mechanistic insight into intravascular coagulation. *Crit Care* 2021;25:95.
29. Jiang WY, Ren J, Zhang XH, et al. CircC3P1 attenuated pro-inflammatory cytokine production and cell apoptosis in acute lung injury induced by sepsis through modulating miR-21. *J Cell Mol Med* 2020;24:11221-9.
30. Li H, Zhang X, Wang P, et al. Knockdown of circ-FANCA alleviates LPS-induced HK2 cell injury via targeting miR-93-5p/OXSRI axis in septic acute kidney injury. *Diabetol Metab Syndr* 2021;13:7.
31. Ma X, Zhu G, Jiao T, et al. Effects of circular RNA Ttc3/miR-148a/Rcan2 axis on inflammation and oxidative stress in rats with acute kidney injury induced by sepsis. *Life Sci* 2021;272:119233.
32. Nan A, Chen L, Zhang N, et al. A novel regulatory network among LncRpa, CircRar1, MiR-671 and apoptotic genes promotes lead-induced neuronal cell apoptosis. *Arch Toxicol* 2017;91:1671-84.
33. Parsons PE, Matthay MA, Ware LB, et al. Elevated plasma levels of soluble TNF receptors are associated with morbidity and mortality in patients with acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 2005;288:L426-31.
34. Lin Q, Liang Q, Qin C, et al. CircANKRD36 Knockdown Suppressed Cell Viability and Migration of LPS-Stimulated RAW264.7 Cells by Sponging MiR-330. *Inflammation* 2021;44:2044-53.
35. Zou Z, Wang Q, Zhou M, et al. Protective effects of P2X7R antagonist in sepsis-induced acute lung injury in mice via regulation of circ_0001679 and circ_0001212 and downstream Pln, Cdh2, and Npr13 expression. *J Gene Med* 2020;22:e3261.
36. Lin H, Wang W, Lee M, et al. Current Status of Septic Cardiomyopathy: Basic Science and Clinical Progress. *Front Pharmacol* 2020;11:210.
37. Yan J, Yang Y, Fan X, et al. Sp1-Mediated circRNA circHipk2 Regulates Myogenesis by Targeting Ribosomal Protein Rpl7. *Genes (Basel)* 2021;12:696.
38. Zhang L, Han B, Liu H, et al. Circular RNA circACSL1 aggravated myocardial inflammation and myocardial injury by sponging miR-8055 and regulating MAPK14 expression. *Cell Death Dis* 2021;12:487.
39. Zhang H, Caudle Y, Shaikh A, et al. Inhibition of microRNA-23b prevents polymicrobial sepsis-induced cardiac dysfunction by modulating TGIF1 and PTEN. *Biomed Pharmacother* 2018;103:869-78.

40. Zhou Y, Qing M, Xu M. Circ-BNIP3L knockdown alleviates LPS-induced renal tubular epithelial cell injury during sepsis-associated acute kidney injury by miR-370-3p/MYD88 axis. *J Bioenerg Biomembr* 2021;53:665-77.
41. Liu J, Liu Y, Zhang L, et al. Down-regulation of circDMNT3B is conducive to intestinal mucosal permeability dysfunction of rats with sepsis via sponging miR-20b-5p. *J Cell Mol Med* 2020;24:6731-40.
42. Hong X, Li S, Wang J, et al. Circular RNA circFADS2 is overexpressed in sepsis and suppresses LPS-induced lung cell apoptosis by inhibiting the maturation of miR-15a-5p. *BMC Immunol* 2021;22:29.
43. Shen W, Zhao X, Li S. Exosomes Derived from ADSCs Attenuate Sepsis-Induced Lung Injury by Delivery of Circ-Fryl and Regulation of the miR-490-3p/SIRT3 Pathway. *Inflammation* 2021;45:331-42.
44. Xiao L, Ma XX, Luo J, et al. Circular RNA CircHIPK3 Promotes Homeostasis of the Intestinal Epithelium by Reducing MicroRNA 29b Function. *Gastroenterology* 2021;161:1303-17.
45. Wei B, Yu L. Circular RNA PRKCI and microRNA-545 relate to sepsis risk, disease severity and 28-day mortality. *Scand J Clin Lab Invest* 2020;80:659-66.
46. Shi X, Ma W, Li Y, et al. CircPRKCI relieves lipopolysaccharide-induced HK2 cell injury by upregulating the expression of miR-545 target gene ZEB2. *Biofactors* 2020;46:475-86.
47. Xiong Y, Wang Y, Tian H, et al. Circ-PRKCI Alleviates Lipopolysaccharide-induced Human Kidney 2 Cell Injury by Regulating miR-106b-5p/GAB1 Axis. *J Cardiovasc Pharmacol* 2021;78:523-33.
48. Li M, Hu J, Peng Y, et al. CircPTK2-miR-181c-5p-HMGB1: a new regulatory pathway for microglia activation and hippocampal neuronal apoptosis induced by sepsis. *Mol Med* 2021;27:45.
49. Xu HP, Ma XY, Yang C. Circular RNA TLK1 Promotes Sepsis-Associated Acute Kidney Injury by Regulating Inflammation and Oxidative Stress Through miR-106a-5p/HMGB1 Axis. *Front Mol Biosci* 2021;8:660269.
50. Li X, Li R, Gong Q, et al. Circular RNA circVMA21 ameliorates lipopolysaccharide (LPS)-induced acute kidney injury by targeting the miR-199a-5p/NRP1 axis in sepsis. *Biochem Biophys Res Commun* 2021;548:174-81.
51. Shi Y, Sun CF, Ge WH, et al. Circular RNA VMA21 ameliorates sepsis-associated acute kidney injury by regulating miR-9-3p/SMG1/inflammation axis and oxidative stress. *J Cell Mol Med* 2020;24:11397-408.
52. Tan M, Bei R. Circ_0091702 serves as a sponge of miR-545-3p to attenuate sepsis-related acute kidney injury by upregulating THBS2. *J Mol Histol* 2021;52:717-28.
53. Zhang X, Dong S. Circ_0091702 relieves lipopolysaccharide (LPS)-induced cell injury by regulating the miR-182/PDE7A axis in sepsis. *Biosci Biotechnol Biochem* 2021;85:1962-70.
54. Tian C, Liu J, Di X, et al. Exosomal hsa_circRNA_104484 and hsa_circRNA_104670 may serve as potential novel biomarkers and therapeutic targets for sepsis. *Sci Rep* 2021;11:14141.
55. Li X, Liu CX, Xue W, et al. Coordinated circRNA Biogenesis and Function with NF90/NF110 in Viral Infection. *Mol Cell* 2017;67:214-27.
56. Jeck WR, Sharpless NE. Detecting and characterizing circular RNAs. *Nat Biotechnol* 2014;32:453-561.
57. Salzman J, Chen RE, Olsen MN, et al. Cell-type specific features of circular RNA expression. *PLoS Genet* 2013;9:e1003777.
58. Liu J, Li D, Luo H, et al. Circular RNAs: The star molecules in cancer. *Mol Aspects Med* 2019;70:141-52.
59. Zhang H, Shen Y, Li Z, et al. The biogenesis and biological functions of circular RNAs and their molecular diagnostic values in cancers. *J Clin Lab Anal* 2020;34:e23049.
60. López-Jiménez E, Rojas AM, Andrés-León E. RNA sequencing and Prediction Tools for Circular RNAs Analysis. *Adv Exp Med Biol* 2018;1087:17-33.
61. Enuka Y, Lauriola M, Feldman ME, et al. Circular RNAs are long-lived and display only minimal early alterations in response to a growth factor. *Nucleic Acids Res* 2016;44:1370-83.